

Enantiopure α -Hydroxy- β -Lactams via Stereoselective Glycosylation¹

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Dedicated to Prof. Usha R. Ghatak

Abstract: Stereospecific glycosylation via the Ferrier rearrangement catalyzed by iodine has been used to obtain both enantiomers of a *trans* 4-aryl-3-hydroxy-2-azetidinone. Unexpectedly, a rhamnol derivative had to be employed for this reaction since a glucal derivative failed to work. This synthesis complements our previously described preparation of the corresponding *cis* isomers by the same methodology. Thus, convenient access has become available to all four homochiral stereoisomers of α -hydroxy- β -lactams some of which are synthons for Taxol[®] and analogs. © 1997 Elsevier Science Ltd.

Recently α -hydroxy- β -lactams have attracted much attention because they are convenient intermediates for the semi-synthesis of the anti-tumor drugs Taxol^R (paclitaxel), Taxotere^R (docletaxel) and their analogs². We³ have shown that variously substituted 3-hydroxy-2-azetidinones are versatile synthons for amino acids, alkaloids, antibiotics and many other types of natural products.

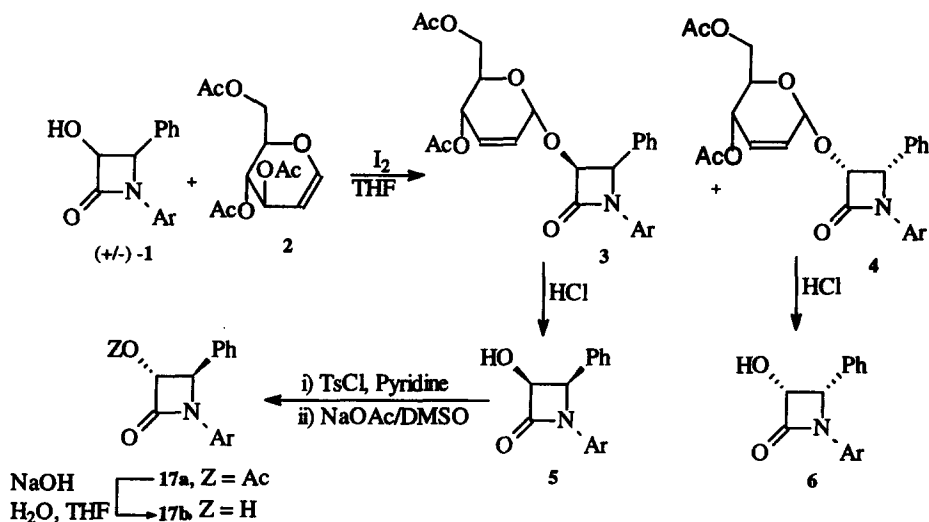
To obtain these end products in optically active forms, we have synthesized enantiopure α -hydroxy- β -lactams by cycloaddition to optically active Schiff bases. Alternatively, we have described a simple method⁴ for stereospecific glycosylation via the Ferrier rearrangement leading to both enantiomers of hydroxy compounds. In particular, we prepared the enantiomeric forms **5** and **6** of *cis* 1-(*p*-anisyl)-3-hydroxy-4-phenyl-2-azetidinone (**1**) by a two step process (Scheme 1):

- (1) the stereospecific formation of two diastereomeric α -glycosides (**3,4**) which were separated by chromatography;
- (2) the hydrolysis of **3** and **4** to optically pure **5** and **6**, respectively. These α -hydroxy- β -lactams are an enantiomeric pair.

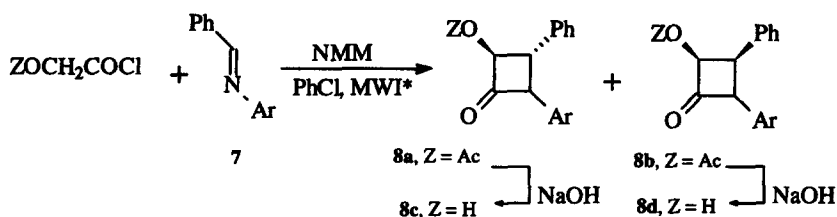
We⁵ have reported recently that easy access to certain *trans* β -lactams can be obtained by conducting the β -lactam ring forming reaction at a high temperature under microwave irradiation⁶ (Scheme 2). Thus, the reaction of the Schiff base **7** with acetoxyacetyl chloride and *N*-methylmorpholine under high power (~800 watts) microwave irradiation produced the *trans* and the *cis* β -lactams **8a** and **8b** in the ratio of 95:5. In contrast, the same annelation reaction under traditional reaction conditions at 0-5°C provided essentially a single isomer - the *cis* α -acetoxy- β -lactam **8b**.

Following our earlier work⁴, we attempted glycosylation of the *trans* β -lactam (**8c**, Z=H) with glucal triacetate **2** in presence of iodine⁷. To our surprise, this attempt was unsuccessful even after prolonging the reaction time to 30h at room temperature. With increased concentration of iodine, there was considerable decomposition of the glucal but no formation of glycosides was observed on monitoring by ¹H NMR spectroscopy and TLC analysis.

Scheme 1



Scheme 2



NMM = N - methylmorpholine; MWI* = microwave irradiation; Ar = p - anisyl

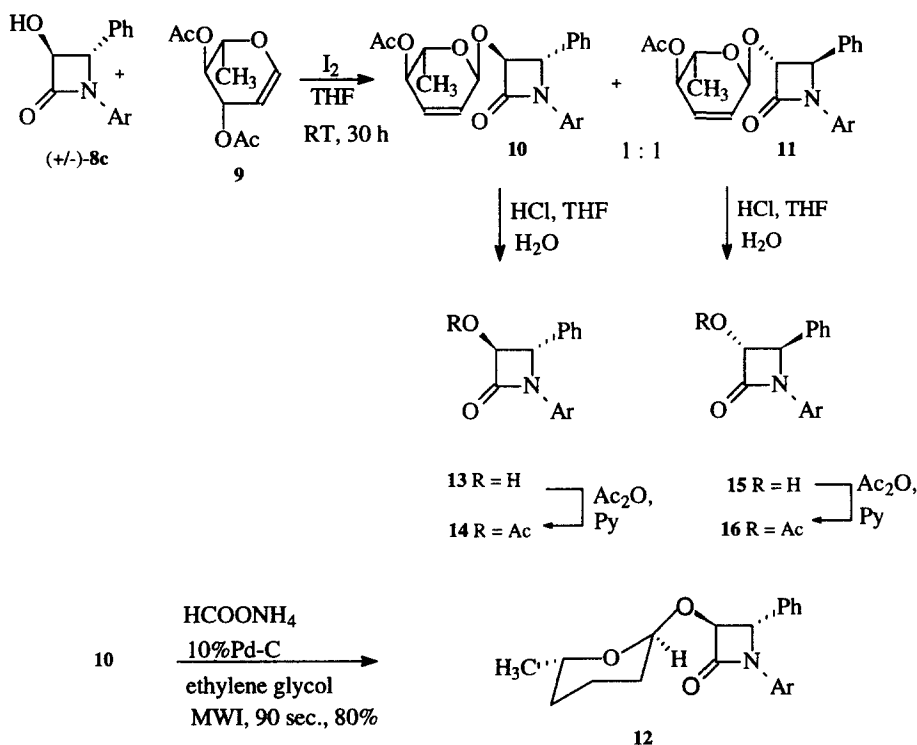
In our previous study we had successfully employed several glycols besides glucal 2. We selected rhamnal diacetate 9 to replace D-glucal triacetate (2). Now the glycosylation of 8c with 9 in the presence of iodine (0.3 eq) proceeded readily and a mixture of two products 10 and 11 in the ratio of 1:1 was obtained in about 50% yield (Scheme 3). These diastereomers (10 and 11) could be separated by column chromatography.

The ^1H NMR spectra of 10 and 11 showed the tell-tale coupling pattern of α -glycosides⁸, no β -glycosides were observed. For obtaining corroborative evidence, 10 was subjected to catalytic reduction⁹ under microwave irradiation as in our earlier studies⁴. The expected reduction product 12¹³ was obtained in which the anomeric proton showed the small couplings characteristic of Jee and Jae and thus supported the α -glycoside structure assigned to 10. It is known that the corresponding β -glycoside would have displayed one small (Jae) and one large (Jaa) coupling¹⁰.

Mild hydrolysis under acidic conditions (HCl/THF/H₂O) of 10 and 11 led to the desired enantiomeric pair 13 and 15 which were converted to their acetates (14 and 16) by room temperature reaction with acetic

anhydride in pyridine solution. Both **14** and **16** were shown to be optically pure and enantiomeric to each other by ^1H NMR studies using an optically active shift reagent as in our earlier work.^{4, 11}

Scheme 3



In our previous study⁴ (Scheme 1) we had shown **5** to have the absolute configuration indicated by its stereostructure by comparison with the known compounds which has been used in other laboratories for the partial synthesis of Taxol^R and Taxotere^R. Tosylation of **5** followed by reaction with sodium acetate had led with $\text{S}_{\text{N}}2$ inversion to a *trans* 3-acetoxy- β -lactam **17a**. The corresponding 3-hydroxy- β -lactam which was readily obtained by the mild hydrolysis of **17a** must therefore have the absolute configuration shown by **17b**.

We observed that **16** was identical with **17a** and thus the absolute configuration for **16**, **11**, and **10** could be assigned with confidence.

With the limited data at hand it is difficult to explain the lack of reaction between glucal triacetate and the *trans* α -hydroxy- β -lactam **8c**. The difference in steric bulk at C-6 between the D-glucal triacetate and L-rhamnal diacetates could be one of the contributing factors.

It is obvious that further work is necessary to obtain a clearer understanding of the iodine catalyzed Ferrier rearrangement and to account for the difference in reactivity of *cis* and *trans* β -lactams. Nonetheless, practical methods are available now, based on the present study, for the preparation of all four stereoisomers of α -hydroxy- β -lactams¹² in the optically pure form. Such synthons, in turn, can lead to diastereomeric forms of Taxol^R and various other natural products.

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References and Notes

1. Studies on Lactams Part 103; for Part 102, see Jayaraman, M.; Manhas, M. S.; Bose, A. K. *Tetrahedron Lett.* **1997**, *38*, 709.
2. For some recent publications on the synthesis of the taxol side chain, see: (a) Hattori, K.; Yamamoto, H. *Tetrahedron Lett.* **1994**, *50*, 2785. (b) Brieva, R.; Crich, J. Z.; Sih, C. *J. Org. Chem.* **1993**, *58*, 1068. (c) Kanazawa, A. M.; Correa, A.; Dennis, J. N.; Luche, M. J.; Green, A. E. *J. Org. Chem.* **1993**, *58*, 255 and references cited therein. (d) Also see, Ojima, I. *Acc. Chem. Res.* **1995**, *28*, 383 and references cited therein.
3. (a) Bose, A. K.; Mathur, C.; Wagle, D. R.; Naqvi, R.; Manhas, M. S. *Heterocycles*, **1995**, 491. (b) Banik, B. K.; Manhas, M. S.; Bose, A. K. *J. Org. Chem.* **1993**, *58*, 307 and references cited therein. (c) also see, Ojima, I. In *The Organic Chemistry of β -Lactams*; Georg, G. I. Ed., VCH Publishers: New York, **1992**; pp. 198-252.
4. Banik B. K.; Manhas, M. S.; Bose, A. K. *J. Org. Chem.* **1994**, *59*, 4714.
5. Bose, A. K.; Banik, B. K.; Manhas, M. S. *Tetrahedron Lett.* **1995**, *36*, 213; also see: Endo, M.; Droghini, R. *BioMed. Chem. Lett.* **1993**, *3*, 2483.
6. For our recent work on Microwave Induced Organic Reaction Enhancement Chemistry, See: Bose, A. K.; Manhas, M. S.; Banik, B. K.; Robb, E. W. *Res. Chem. Intermed.* **1994**, *20*, 1; also see: Bose, A. K.; Jayaraman, M.; Okawa, A.; Bari, S. S.; Robb, E. W.; Manhas, M. S. *Tetrahedron Lett.* **1996**, *39*, 6989; Banik, B. K.; Manhas, M. S.; Robb, E. W.; Bose, A. K. *Heterocycles*, **1997**, *44*, 405 and references cited therein.
7. For iodine catalyzed glycosylation reaction via Ferrier rearrangement, see: Koreeda, M.; Houston, T. A.; Shull, B. K.; Klemke, E.; Tuinman, R. J. *Synlett*, **1995**, 90.
8. Ferrier, R. J. *Adv. Carbohydr. Chem. Biochem.* **1969**, 199 and references cited therein.
9. The reduction of the olefinic group in **10** was conducted by catalytic transfer hydrogenation under microwave irradiation, see: Bose, A. K.; Banik, B. K.; Barakat, K. J.; Manhas, M. S. *Synlett*, **1993**, 575.
10. The ^1H NMR spectrum of the hydrogenated product **12** obtained from **10** indicated the α -glycosidic linkage. See: Lemieux, R. U.; Stevens, J. D. *Can. J. Chem.* **1965**, *43*, 2059.
11. Compound **13**: mp 152°C; $[\alpha]_{\text{D}}^{23} + 110^\circ$ (c 1.0, CH_3OH). Compound **15**: mp 152°C; $[\alpha]_{\text{D}}^{23} - 108^\circ$ (c 1.0, CH_3OH). ^1H NMR of **14** and **16** in the presence of a chiral shift reagent confirmed the absolute configuration of these β -lactams and their mirror image relationship.
12. All new compounds described here gave satisfactory spectral and elemental analyses.
13. Data for **12**: IR (CH_2Cl_2): 1740, 1610 cm^{-1} ; ^1H NMR (200MHz, CDCl_3) δ : 7.35(s, 5H), 7.25(d, $J=6.90\text{Hz}$, 2H), 5.13 (brs, 1H), 4.89(d, $J=1.51\text{Hz}$, 1H), 4.65(d, $J=1.55\text{Hz}$, 1H), 4.13-3.94(m, 1H), 3.73(s, 3H), 2.05-1.26(m, 6H), 1.07(d, $J=6.29\text{Hz}$, 3H); ^{13}C NMR: 164.09, 156.35, 148.22, 136.52, 130.88, 129.01, 128.50, 126.15, 116.90, 114.36, 110.49, 99.26, 88.17, 84.62, 65.92, 64.48, 55.37, 32.75, 29.07, 21.59, 21.03, 17.22, 9.13. Anal. Calcd for $\text{C}_{22}\text{H}_{25}\text{O}_4\text{N}$: C, 71.91; H, 6.85; N, 3.81. Found: C, 71.80; H, 6.73; N, 3.64.

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